Antifungal and Antibacterial Properties of Graphene-based Nanomaterials: A Mini-review

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ABSTRACT

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In recent years, the availability and use of various antibiotics and antimicrobial agents have resulted in increase of drug resistant pathogens. Therefore, scientist's attention has been diverted to find a suitable replacement for antimicrobial treatment. Graphene (G), as a twodimensional (2D) carbon-based nanomaterials (CBNs) has a unique physicochemical properties including thermal, optical and electrically conductive activities along with high surface-to-volume ratio and mechanical strength. Also, they have gained significant attention in biomedical application, such as regenerative medicine and drug delivery carriers. With the emergent of nanomaterials over the past decades, there has been a growing interest in using GBNs to develop new antifungal and antimicrobial nanomaterials due to their diverse antifungal and bactericidal mechanisms and relatively low cytotoxicity towards mammalian cells. Numerous reviews on GBNs are available with different perspective. In this review, we summarized the latest progresses towards an understanding of the antifungal and antibacterial properties of GBNs for developing a new antifungal and antibacterial materials

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INTRODUCTION

Currently, the prevalence of antibiotic-resistant bacteria and costs of treatment has become public health concern [1]. Microorganisms (e.g. fungi and bacteria) can easily attach to the surfaces of medical apparatus and colonize on their surfaces. These contaminations are threat to human health and might lead to economic losses [2]. The advances in the field of nanotechnology has created good opportunities to solve these issues. Consequently, synthesis and application of new material to overcome these threats are highly desirable. In recent years, Nano materials have paved the way to create novel antimicrobial agents with exclusive chemical and physical properties [3- 5]. Many nanomaterials including silver (Ag) [6, 7],

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zinc oxide (ZnO) [8], titanium dioxide (TiO₂) [9], iron oxide (Fe₃O₄) [10], copper oxide (CuO) [11], magnesium oxide (MgO) [12], nitric oxide (NO) nanoparticles[13] have been shown to have good antibacterial properties. Thus, expansion of such nanoparticles to combat microbial agents can be an important component for the decontamination process in the near future.

Another kind of nanomaterials that has been investigated widely, are carbon-based nanomaterials. Graphene-based nanomaterials (GBNs) as an important member of this family has been investigated in last decade due to its unique properties, such as large surface-to-volume ratio, mechanical flexibility and thermal stability [14- 17]. GBNs are a promising candidate in polymeric

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Fig. 1. Graphene (G) and Geraphene oxide (GO) structures [44].

nanocomposite synthesis [18, 19], catalysis [20- 23], solar cells [24], biosensors [25], drug delivery and gene delivery [26, 27], imaging [28-30], photothermal therapy [31, 32], tissue engineering [33, 34], and stem cell technology [35, 36].

Moreover, GBNs have established to have antibacterial activities. Graphene oxide (GO) and reduced graphene oxide (rGO) are toxic to both Gram-positive and Gram-negative bacteria [37, 38]. The antifungal and antibacterial applications of GBNs are still relatively novel. In the last decade attention to GBNs has increased exponentially. Numerous reviews on GBNs are available with different perspective [39-42]. In this review, we summarized the latest progresses towards an understanding of the antifungal and antibacterial properties of GBNs. In the first section, we introduced GBNs, and the approaches to their fabrication. The second part briefly looks at their main antifungal and antibacterial mechanism. The last part include several examples of GBNs application as an antifungal and antibacterial nanomaterial.

The GBN family and their fabrication

GBNs can be defined and classified according to their features including morphology and composition, the average of lateral dimension and the number of G layers, which is determined by the atomic carbon/oxygen ratio, material's size and degree of deformability, thickness, specific surface area, and the bending elasticity of the material properties [43] [44]. The morphological and compositional based classification is essential when working with GBNs. Depending on the synthetic methods, G can be prepared in various morphologies for example sheets, platelets, ribbons and quantum dots (QDs). According to this family of nanomaterials, there are various nanostructures including graphene (G), graphene

oxide (GO), reduced graphene (rGO), ultrafine graphite that are between 5 to 10 sheets and below 100 nm in thickness, as well as graphene ribbons, graphene quantum dots (GQDs), and pristine graphene (pG) [45-48]. G is a single monolayer of sp²-bonded carbon with a single atom thick of 2D-model graphite carbon material, which can be fabricated from graphite by scotchtape technique,[49] chemical exfoliation [50], chemical vapor deposition [51], arc discharge [52], and decomposition of carbide phases [53]. GO structure consists of single-atom-thick carbon sheets with carboxylate groups on the periphery, where they provide pH dependent negative surface charge and colloidal stability. GO is a single layer that can be produced via reaction of crystalline graphite with a mixture of oxidizing agents and sonication or other dispersion methods, (Fig. 1) shows G and GO [44].

Also, it consists of epoxy, hydroxyl and carboxylic acid groups on its surface and edges. rGO is a single layer that can be fabricated from GO through under reducing condition, consisting of high-temperature thermal treatment and chemical treatments with hydrazine (N_2H_4) or other reducing agents [54]. GQDs are small pieces of G with a 2D lateral size less than 100 nm, which have been synthesized by oxidative cleavage [55], hydrothermal or solvothermal method [56-58], macrowave-assisted / ultrasonic-assisted process [59], electrochemical oxidation [60], and carbonization [61]. pG is an apolar and hydrophobic member of GBNs that its dispersibility in aqueous media is improved by oxidizing. Due to unique arrangement of sp^2 bonded carbon atoms, each GBNs can exhibit remarkable different physical, morphological and chemical properties (Fig. 2)

GBNS antifungal and antibacterial mechanisms The antimicrobial mechanisms responsible for

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GBN Transcriptional Protein **Dysfunction** Arres

Fig. 3. Different possible mechanisms of antibacterial activity of GBNs

G and GO and other GBNs have been investigated widely. The most common proposed mechanisms are oxidative stress induction, protein dysfunction, membrane damage, and transcriptional arrest (Fig. 3) [62-66]. Generating reactive oxygen species (ROS) is the main reason for nanomaterial toxicity [45, 67-69]. Some antioxidant enzymes, such as glutathione peroxidase or superoxide dismutase, are able to diminish and eliminate ROS generation. By disrupting these enzymes balance, proteins, deoxyribonucleic acid (DNA), and lipids can be damaged. In addition, GO and rGO nanosheets have shown Fenton-like catalytic activity [70, 71]. Hence, there is a structural connection between GBNs and their redox activity that supports the ROS generation ability of GBNs as an antimicrobial mechanisms. Microorganism membrane damage is another possible outcome of hydrophobic interaction between GBNs and the membrane phospholipids that correlate with size of GBNs [72-74]. Although the protein dysfunction and

transcriptional arrest were not typically proposed to be the primary antibacterial mechanism of GBNs, sometimes they contribute to the antibacterial activities. Investigations showed that G-Fe₃O₄ leads to *E. coli* protein aggregation, while, the Fe₃O₄ causes less protein degradation by itself, the same as tungsten oxide (WO_3) nanoparticles by itself [63, 64]. These results show that protein dysfunction can be augmented by G structure in comparison to when they are alone. Due to $π$ -π stacking interactions, GBNs can interact with DNA in several groups. For instance, the presence of GO alongside $Cu²⁺$ can affect DNA cleavage by the chelation of $Cu²⁺$ ions to oxygen functional groups on the GO nanosheets [75]. Larger GO nanosheets show significant reduction in *E. coli* viability assay (40 μ g/mL, 2 h) in comparison to smaller nanosheets [76].

The antifungal activity of GBNs

Fungi are able to easily colonize the surfaces of most materials and devices, and they can quickly spread fungal spores. Human health can be threatened by the formation of fungal contamination that might lead to vast economic losses. Consequently, suitable material against fungi are extremely desired. GBNs antifungal activity was studied by synthesizing different types of GBNs, especially G, GO, and rGO.

The antifungal activity of RGO nanosheets

Antifungal activity of rGO against pathogenic fungi can be used to develop GBNs as a broad spectrum antifungal agents. As a breakthrough, the antifungal activity of rGO (0-500 μg/mL) Sawangphruk et al. [77], studied against three fungal species of *A. niger, A. oryzae,* and *F. oxysporum*, and showed the efficacy of rGO against fungi. The half maximal inhibitory concentration (IC₅₀) values of the rGO against *F. oxysporum, A. niger* and *A. oryzae* were 50, 100, and 100 μg/mL, respectively. The probable antifungal mechanism, is the interaction of rGO nanosheets with the cell walls of fungi. After that the ROS generation of rGO nanosheets was able to chemically react with the organic functional groups of chitin and other polysaccharides on the cell walls of fungi and induce antifungal activity.

The antifungal activity of GO nanosheets

The main antifungal activity of GO is related to its sharp edges that can cause plasma membrane stress on pathogenic cells. To understand the interaction mechanism of GO, an investigation was performed by Chen et al., [72] on bacterial and fungal pathogens, such as *P. syringae* and *X. campestris pv. Undulosa, F. graminearum,* and *F. oxysporum*. The results showed that GO inhibits nearly 90% of the bacteria and repress 80% macroconidia germination along with partial cell swelling and lysing. The proposed mechanism for the toxicity of GO against both the bacterial and the fungal pathogens was a wide range of GO nanosheets aggregation, which resulted in local perturbation of cell membrane, reduced cell membrane potential, and electrolytes leakage. Moreover, due to the high efficiency of GO for photothermal treatment in the near-infrared (NIR (region, it can be an effective photothermal material. The photothermal treatment of GO was investigated by Khan et al., [78], for antifungal activity to avoid wound healing infection. As a non-invasive and cheap alternative method, this therapy showed remarkable healing property for infected wounds on the dorsal surface of mice (Fig. 4). The antifungal activity of GO on *S. cerevisiae* and *C. utilis* was investigated. Results showed that

the laser mediated surface activation of GO causes great antifungal efficiency (Fig. 5)

In a study by Zhu et al., [79] selected *S. cerevisiae* as a model and the potential toxicity of GO was evaluated at the concentration ranging from 0 to 600 mg/mL. The results showed a dose dependent cytotoxicity. The antifungal mechanism was attributed to the synergy of reduced mitochondrial transmembrane potential and increased ROS generation. Hence, the expressions of apoptosis-related genes, such as SOD, Yca1, Nma111 and Nuc1 were significantly changed. Xie et al., [80] exposed *P. chrysosporium*, white rot fungus to GO at the concentration of 0-4 mg/mL for 7 day. Their results showed that low concentrations of GO stimulate the cells growth and causes more acidic pH values of the culture media. In addition, the scanning electron microscopy investigations images exhibited that GO induce the disruption of fiber structure of *P. chrysosporium*, where some very long and thick fibers were formed at 4 mg/mL. In another study, single-walled carbon nanotubes (SWCNTs), multi-walled carbon nanotubes (MWCNTs), GO, rGO, fullerene (C60) and activated carbon (AC)

Fig. 4. Standard microdilution protocol of the antibacterial activities of GO and their photothermal treatment [78].

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Fig. 5. (a) Location of wounds (CW, LW, and GOLW) on mice model, (b) three fresh wounds mounted on mice dorsal surface, (c) show infection started after 3 days of injected S. aureus on wounds, (d) the picture shows the condition of wound after 6 days of treatment on the wound, (e) the photographs also shows the condition of wound after 9 days of treatment on the wound, and (f) show changed in the treatment of wounds on the dorsal surface of mice model by using GO and Nd-YAG laser [78].

Fig. 6. Inhibition zone of GO, GO-AgNPs and CNSs-AgNPs to (a) C. albicans and (b) C. tropical by the disk diffusion assay [84].

were examined by Wang et al., [81] against two important plant pathogenic fungi, *F. graminearum* and *F. poae*. The strongest antifungal activity was observed for SWCNTs, followed by MWCNTs, GO, and rGO, where C60 and AC exhibited no noteworthy antifungal activity. The antifungal mechanism included three steps; depositing on

the surface of the spores, preventing water uptake and prompting plasmolysis.

The antifungal activity of GO nanocomposites

The synergistic effect of GO and other nanoparticles can be applied to formulate more efficient antimicrobial products [82, 83].

Hence, in recent years scientists have explored Ag nanocomposites more than ever before. To increase antifungal activity of carbon nano scrolls (CNSs), Li et al., [84] filled it with silver nanoparticles (AgNPs) and compared it with antifungal activity of GO-AgNPs nanocomposite. The CNSs-AgNPs exhibited prolonged activity against *C. albicans* and *C. tropical* in comparison with the GO-AgNPs nanocomposites. The results of antifungal activities of GO and its AgNPs nanocomposites showed that there were no inhibition zone for GO; while for GO-AgNPs samples a clear inhibition zones were observed. By increasing incubation time, the inhabitation zone become smaller, and the viable fungal colony increased (Fig. 6)*.* Moreover, the inhibition zone of *C. albicans* was much smaller than the *C. tropical* in the same concentration of GO-AgNPs and same incubation time.

In another study by Chen et al., [85] they explored antifungal activity of GO-AgNPs nanocomposite against phytopathogen *F. graminearum in vitro* and *in vivo*. The GO-AgNPs nanocomposite showed to be three to seventimes more potent than pure AgNPs and GO, respectively. The antifungal mechanism was based on the notable synergistic effect of GO-AgNPs, making physical injury and generating chemical reactive oxygen species. In addition to silver nanocomposites, G-TiO₂ nanocomposite was investigated by Karimi et al., [86] as a new route to

prepare antibacterial and antifungal cotton fabric without toxicity. The result showed that G-TiO₂ nanocomposite-coated cotton has an excellent antibacterial and antifungal activity on bacteria (*E. coli and S. aureus*) and fungi (*C. albicans*). Indeed, G was added to TiO₂ nanoparticles aiming to facilitate effective bacterial decomposition by increasing the contact between nanoparticles and microorganisms. Furthermore, fabrics treated with GO did not show any antimicrobial activity. Graphene oxide-borneol (GOB) composite, is borneol-grafting with great antifungal effect on *M. racemosus*. In comparison with GO nanosheet, GOB composite displayed no significant antifungal activity. In addition to long-term antifungal effect of GOB composite, the fallen spore does not germinate even after 5 days [87]. The studies are summarized in the Table 1.

The antibacterial activity of GBNs

World Health Organization (WHO) reports showed that in recent years, death of millions of people has been due to the diseases created by bacterial infections [88]. Thus, the treatment of bacterial diseases using antimicrobial drugs are vital. Also, there is an urgent need for novel and effective antimicrobial agents to fight against the bacterial infections [89, 90]. G has been found to be a capable candidate as an antibacterial material due to its bacterial toxicity. Therefore, the toxicity investigation of GBNs in microorganisms as a

Table 1. The antifungal activity of GBNs

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new class of antibacterial material, is vital for their production in environmental and clinical applications [37].

The antibacterial activity of RGO and GO nanosheets

For the first time Hu et al., [37] investigated antibacterial properties of GBNs by studying the interaction of Gram-negative bacteria, *E. coli* DH_{ϵ} with GO. The results showed that GO at a concentration of 85 μg/mL could significantly suppress the growth of *E. coli,* while having low cytotoxicity for mammalian cells. Transmission electron microscope (TEM) analysis showed that antibacterial properties were attributed to damage cell membrane, which results in leakage of the cytoplasm (Fig. 7). They further found that macroscopic GO papers prepared by vacuum filtration of the GO suspension could effectively restrain the growth of *E. coli*.

Also, dental caries and periodontal diseases are related to microbes, such as S. mutans, P. gingivalis and F. nucleatum. The antibacterial influence of GO was examined by He et al., [91]. TEM analysis showed GO can disrupt the cell walls, membrane integrity and leakage of the intracellular contents. The antibacterial activity were also observed for a UV irradiated GO by Veerapandian et al., [92]

that showed higher antibacterial activity for UV irradiated GO due to more cell disruption action than typical GO nanosheets. The antibacterial activity of GO and rGO was evaluated by Gurunathan et al., [93] using cell viability, ROS generation and DNA fragmentation assays. The results suggest that GO and rGO possessed a time and concentration dependent antimicrobial activity against *E. coli.* In comparison with rGO, GO formed more superoxide anions than rGO. Therefore, the bacterial cell death might be due to oxidative stress that consequently leads to DNA fragmentation. The antibacterial activity of pG, GO, and rGO against food-borne bacterial pathogens, such as *L. monocytogenes* and *S. enterica* were evaluated by Kurantowicz et al., [94]. This study reported GO to have the highest antibacterial activity due to bacteria adherence at the surface of GO, while with pG and rGO, they adhered to their edges. Also, Wu et al.,[95] observed a concentration dependent antibacterial activity for GO against K. pneumonae.

Some reports showed that GO exhibited no significant antimicrobial effect against *E. coli* or *P. aeruginosa* bacteria alone; however, Ag nanoparticle-modified GO could effectively inhibit bacterial growth. Interestingly, another research showed that GO presented neither intrinsic

Fig. 7. Antibacterial activity of GO. (a) Metabolic activity of E. coli incubation with 20 and 85 mg/mL of GO at 37 °C for 2 h. (b) Antibacterial activity of 85 mg/mL GO against E. coli DH5 cells. (c,d) TEM images of untreated E. coli (c) and E. coli exposed to GO nanosheets (d) at 37 ° C for 2 h [37].

GBNs	Treatment concentration	Fungal	Summary results	Reference
rGO	0 and 500 μ g/mL	A. niger, A. oryzae, F. oxysporum	rGO was effective against on the nonpathogenic A. oryzae and on the pathogenic A , niger and F , oxysporum	$[77]$
GO	$500 \mu g/mL$	F . graminearum, F. oxysporum	nearly 90% of the bacteria and repressed 80% macroconidia germination along with partial cell swelling and lysis	$[72]$
SWCNTs. MWCNT, GO, rGO, C60, AC	0 and 500 μ g/mL	F. graminearum, F. poae	No noteworthy antifungal activity was detected for C60 and AC	[81]
GO	0 to 600 μ g/mL	S. cerevisiae	Dose dependent cytotoxicity	[79]
GO	$0-4$ mg/mL	P. chrysosporium	Induced the disruption of the fiber structure	[80]
$GO-Ag$	$0-8 \mu$ g/mL	C. albicans. C. tropical	CNSs-AgNPs exhibited ideal lengthened activities against Candida albicans and Candida tropical compared with the GO-AgNPs nanocomposites based on silver nanoparticles directly deposited on the surface of grapheme oxides, which is caused by CNS-AgNPs' controlled durative slow-releasing of silver ion	$[84]$
GO-AgNPs		F. graminearum	Exhibited nearly a three and seven-fold increase of inhibition efficiency over pure AgNPs and GO making physical injury and chemical reactive oxygen species generation	[85]
GOB		M. racemosus	It is also worth noting that on the GOB sample the fallen spore does not germinate even after 5 days	$[87]$
$G-TiO2$		C. albicans	They had negligible toxicity and possessed excellent antimicrobial activity	[86]

Table 2. The antibacterial activity of GBNs

antibacterial functions nor cytotoxicity properties to mammalian cells [96, 97]. Several studies evaluate the antibacterial activity of Gram-negative and Gram-positive bacteria and showed that GO inhibited Gram-positive bacteria more effectively than Gram-negative bacteria, while some Gramnegative bacteria, such as *E. coli*, were resistant to GO [98]. *E. coli*, gram-negative and *S. aureus*, gram-positive were selected by Akhavan et al., [38] as model bacteria to investigate the bacterial toxicity of GO and rGO nanowalls. Results showed bacteria interaction between the very sharp edges of the nanowalls causes cell membrane damage. In addition, the cell membrane of Gram-positive *S. aureus* was strictly damaged in comparison to the Gram-negative *E. coli,* which was due to sharp edges and better charge-transfer ability of rGO nanowalls and bacteria. Moreover, rGO nanowalls exhibited stronger antibacterial activity than GO nanowalls. The antibacterial mechanism of G was also studied by exploring the interactions between different types of GBNs with the Gramnegative *E. coli*. Again, GO showed the strongest antibacterial activity under similar concentrations and incubation time among all materials, followed by rGO, graphite, and graphite oxide. Their antibacterial mechanisms were attributed to the synergy of the membrane stress and oxidative stress induced by the interactions between bacteria and materials. However, by increasing the concentration of GBNs, the inhibition activity

against the growth of Gram-negative *E. coli* and Gram-positive *B. subtilis* was increased [99]. A similar study by Chen et al., [100] studied the antibacterial activity of rGO and GO. And, GO showed higher bactericidal effects due to its sharp edges and production of ROS.

Furthermore, disrupting the membrane integrity, ROS generation can be potentially made by antibacterial activity. Krishnamoorthy et al., [101] examined antibacterial activity of G nanosheets that can be applied in the development of biomedical devices. They observed that G nanosheets have antibacterial activities against *E. coli, S. typhimurium, E. faecalis*, and *B. subtilis*. These results support the idea of G, as a hopeful antibacterial material with low mammalian cell cytotoxicity. Conversely, recent studies have shown that GO might miss any antibacterial properties [96]. These studies are summarized in Table 2.

The antibacterial activity of GBNs nanocomposites

The advancement of nanotechnology provides opportunities to prepare antibacterial G nanocomposits. Antibacterial properties of GBNs include ZnO/GO [102], TiO₂/GO [103], Ti-GO-Ag [104], and CuO/rGO nanocomposites [105] have been explored recently. To date, Ag nanoparticlemodified GO films, not bare GO, have exhibited stronger antibacterial activity. The oxygencontaining functional group of GO adhere to lipopolysaccharides of bacteria through hydrogen

bonds formation between the lipopolysaccharides of the bacteria and the oxygen-containing functional groups of GO [106-108]. Hence, GO decreases the intake of nutrition from the media and increases the interaction between Ag nanoparticles and bacteria [109]. Ag can also disrupt the bacterial membrane that prevents the respiration and replication of bacteria, which leads to cell death [110]. The Ag-modified GO nanostructure exerts its antibacterial effect through a "capturing-killing process" that increases the deposition of bacteria, as well as the contact between cells and Ag-modified GO nanoparticles [96]. Due to these controversial findings, further studies should be carried out to determine the detailed mechanisms and controlling factors with respect to the interaction between GBNs and microbes. There are some studies that have shown recent progress in antimicrobial activity of G nanocomposits [111, 112].

CONCLUSION

Today antibiotic resistance has emerged as a strong health concern worldwide. To eradicate this problem, synthesis and application of new antimicrobial materials are necessary and required. Emerging nanotechnology has provided a suitable platform to resolve the problem of resistance, by the use of antimicrobial nanomaterials, identified as nanomaterials with antibacterial properties to contest infections by antibiotic-resistant bacteria. Several investigations have focused on the antimicrobial mechanism of GBNS, but a deeper and more consistent understanding of the underlying molecular mechanisms is required. Recent studies revealed that GBNs have high efficiency in antifungal and antibacterial activity via damage of cell membranes and other mechanisms. To increase GBNs antimicrobial effect, nanocomposite preparation by the incorporation of inorganic nanostructures has increased their antimicrobial properties. GBNs preferably as a new group of nanomaterials can be used in nanomedicine in the near future.

CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest regarding the publication of this manuscript.

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REFERENCES

- 1. Moritz M, Geszke-Moritz M. The newest achievements in synthesis, immobilization and practical applications of antibacterial nanoparticles. Chem. Eng. J., 2013; 228: 596- 613.
- 2. Yu Q, Wu Z, Chen H. Dual-function antibacterial surfaces for biomedical applications. Acta biomater., 2015; 16: 1-13.
- 3. Zhu X, Radovic-Moreno AF, Wu J, Langer R, Shi J. Nanomedicine in the management of microbial infection– overview and perspectives. Nano today, 2014; 9(4): 478- 498.
- 4. Franci G, Falanga A, Galdiero S, Palomba L, Rai M, Morelli G, et al. Silver nanoparticles as potential antibacterial agents. Molecules, 2015; 20(5): 8856-8874.
- 5. Ebrahiminezhad A, Taghizadeh S, Berenjian A, Heidaryan Naeini F, Ghasemi Y. Green synthesis of silver nanoparticles capped with natural carbohydrates using ephedra intermedia. Nanosci. Nanotechnol., 2017; 7(1): 104-112.
- 6. Wong KK, Liu X. Silver nanoparticles—the real "silver bullet" in clinical medicine? Med. Chem. Comm., 2010; 1(2): 125-131.
- 7. Mallakpour S, Abdolmaleki A, Borandeh S, Sabzalian MR. One pot fabrication of optically active and efficient antibacterial poly(amide-benzimidazole-imide)/Ag bionanocomposite. J. Polym. Res., 2015; 22(7): 129.
- 8. Gordon T, Perlstein B, Houbara O, Felner I, Banin E, Margel S. Synthesis and characterization of zinc/iron oxide composite nanoparticles and their antibacterial properties. Colloids Surf., A, 2011; 374(1-3): 1-8.
- 9. Srisitthiratkul C, Pongsorrarith V, Intasanta N. The potential use of nanosilver-decorated titanium dioxide nanofibers for toxin decomposition with antimicrobial and selfcleaning properties. Appl. Surf. Sci., 2011; 257(21): 8850- 8856.
- 10. Kanagasubbulakshmi S, Kadirvelu K. Green synthesis of Iron oxide nanoparticles using Lagenaria siceraria and evaluation of its Antimicrobial activity. Defence Life Sci. J., 2017; 2(4): 422-427.
- 11. Jadhav MS, Kulkarni S, Raikar P, Barretto DA, Vootla SK, Raikar U. Green biosynthesis of CuO & Ag–CuO nanoparticles from Malus domestica leaf extract and evaluation of antibacterial, antioxidant and DNA cleavage activities. New J. Chem., 2018; 42(1): 204-213.
- 12. Karthik K, Dhanuskodi S, Kumar SP, Gobinath C, Sivaramakrishnan S. Microwave assisted green synthesis of MgO nanorods and their antibacterial and anti-breast cancer activities. Mater. Lett., 2017; 206: 217-220.
- 13. Martinez LR, Han G, Chacko M, Mihu MR, Jacobson M, Gialanella P, et al. Antimicrobial and healing efficacy of sustained release nitric oxide nanoparticles against Staphylococcus aureus skin infection. J. Invest. Dermatol., 2009; 129(10): 2463-2469.

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- 14. Balandin AA, Ghosh S, Bao W, Calizo I, Teweldebrhan D, Miao F, et al. Superior thermal conductivity of single-layer graphene. Nano Lett., 2008; 8(3): 902-907.
- 15. Novoselov KS, Geim AK, Morozov S, Jiang D, Katsnelson M, Grigorieva I, et al. Two-dimensional gas of massless Dirac fermions in graphene. Nature, 2005; 438(7065): 197-200.
- 16. Zhu Y, Murali S, Cai W, Li X, Suk JW, Potts JR, et al. Graphene and graphene oxide: synthesis, properties, and applications. Adv. Mater., 2010; 22(35): 3906-3924.
- 17. Mallakpour S, Abdolmaleki A, Borandeh S. Covalently functionalized graphene sheets with biocompatible natural amino acids. Appl. Surf. Sci., 2014; 307: 533-542.
- 18. Abdolmaleki A, Mallakpour S, Borandeh S. Improving interfacial interaction of l‐phenylalanine‐functionalized graphene nanofiller and poly(vinyl alcohol) nanocomposites for obtaining significant membrane properties: Morphology, thermal, and mechanical studies. Polym. Compos., 2016; 37(6): 1924-1935.
- 19. Mallakpour S, Abdolmaleki A, khalesi Z, Borandeh S. Surface functionalization of GO, preparation and characterization of PVA/TRIS-GO nanocomposites. Polymer, 2015; 81: 140- 150.
- 20. Wu G, Xu P, Guo Z, Chen Y, Lu NL. Graphene Composite Catalysts for Electrochemical Energy Conversion. Multifunctional Nanocomposites for Energy and Environmental Applications, 2018: 203-230.
- 21. Ren X, Guo H, Feng J, Si P, Zhang L, Ci L. Synergic mechanism of adsorption and metal-free catalysis for phenol degradation by N-doped graphene aerogel. Chemosphere, 2018; 191: 389-399.
- 22. Yin R, Guo W, Du J, Zhou X, Zheng H, Wu Q, et al. Heteroatoms doped graphene for catalytic ozonation of sulfamethoxazole by metal-free catalysis: Performances and mechanisms. Chem. Eng. J., 2017; 317: 632-639.
- 23. Keshipour S, Kulaei M, Ahour F. Graphene Oxide Nano-Sheets-Supported Co (II)-d-Penicillamine as a Green and Highly Selective Catalyst for Epoxidation of Styrene. Iran. J. Sci. Technol., A, 2017; 1-10.
- 24. Taki M, Rezaei B, Fani N, Borandeh S, Abdolmaleki A, Ensafi AA. Beneficial effects of amino acid-functionalized graphene nanosheets incorporated in the photoanode material of dye-sensitized solar cells: A practical and theoretical study. Appl. Surf. Sci., 2017; 403: 218-229.
- 25. Saini D. Synthesis and functionalization of graphene and application in electrochemical biosensing. Nanotechnol. Rev., 2016; 5(4): 393.
- 26. Yao X, Niu X, Ma K, Huang P, Grothe J, Kaskel S, et al. Graphene Quantum Dots‐Capped Magnetic Mesoporous Silica Nanoparticles as a Multifunctional Platform for Controlled Drug Delivery, Magnetic Hyperthermia, and Photothermal Therapy. Small, 2017; 13(2): 1602225.
- 27. Foreman H-CC, Lalwani G, Kalra J, Krug LT, Sitharaman B. Gene delivery to mammalian cells using a graphene nanoribbon platform. J. Mater. Chem., B, 2017; 5(12): 2347-2354.
- 28. Cheng R, Peng Y, Ge C, Bu Y, Liu H, Huang H, et al. A turnon fluorescent lysine nanoprobe based on the use of the Alizarin Red aluminum (III) complex conjugated to graphene oxide, and its application to cellular imaging of lysine. Microchim. Acta, 2017; 184(9): 3521-3528.
- 29. Yoo JM, Kang JH, Hong BH. Graphene-based nanomaterials for versatile imaging studies. Chem. Soc. Rev., 2015; 44(14): 4835-4852.
- 30. Lundeberg MB, Gao Y, Woessner A, Tan C, Alonso-González P, Watanabe K, et al. Thermoelectric detection and imaging of propagating graphene plasmons. Nat. mater., 2017; 16(2): 204–207.
- 31. Robinson JT, Tabakman SM, Liang Y, Wang H, Sanchez Casalongue H, Vinh D, et al. Ultrasmall reduced graphene oxide with high near-infrared absorbance for photothermal therapy. J. Am. Chem. Soc., 2011; 133(17): 6825-6831.
- 32. Tayyebi A, Akhavan O, Lee B-K, Outokesh M. Supercritical water in top-down formation of tunable-sized graphene quantum dots applicable in effective photothermal treatments of tissues. Carbon, 2018; 130: 267-272.
- 33. Nie W, Peng C, Zhou X, Chen L, Wang W, Zhang Y, et al. Three-dimensional porous scaffold by self-assembly of reduced graphene oxide and nano-hydroxyapatite composites for bone tissue engineering. Carbon, 2017; 116: 325-337.
- 34. Bai RG, Ninan N, Muthoosamy K, Manickam S. Graphene: A versatile platform for nanotheranostics and tissue engineering. Prog. Mater. Sci., 2018; 91: 24-69.
- 35. Suryaprakash S, Li M, Lao Y-H, Wang H-X, Leong KW. Graphene oxide cellular patches for mesenchymal stem cell-based cancer therapy. Carbon, 2018; 129: 863-868.
- 36. Shah S, Yin PT, Uehara TM, Chueng STD, Yang L, Lee KB. Guiding Stem Cell Differentiation into Oligodendrocytes Using Graphene‐Nanofiber Hybrid Scaffolds. Adv. Mater., 2014; 26(22): 3673-3680.
- 37. Hu W, Peng C, Luo W, Lv M, Li X, Li D, et al. Graphene-based antibacterial paper. ACS Nano, 2010; 4(7): 4317-4323.
- 38. Akhavan O, Ghaderi E. Toxicity of graphene and graphene oxide nanowalls against bacteria. ACS Nano, 2010; 4(10): 5731-5736.
- 39. Zhang Q, Wu Z, Li N, Pu Y, Wang B, Zhang T, et al. Advanced review of graphene-based nanomaterials in drug delivery systems: Synthesis, modification, toxicity and application. Mater. Sci. Eng., C, 2017; 77: 1363-1375.
- 40. Lin J, Chen X, Huang P. Graphene-based nanomaterials for bioimaging. Adv. Drug Delivery Rev., 2016; 105: 242-254.
- 41. Goenka S, Sant V, Sant S. Graphene-based nanomaterials for drug delivery and tissue engineering. J. Controlled Release, 2014; 173: 75-88.
- 42. Tonelli FM, Goulart VA, Gomes KN, Ladeira MS, Santos AK, Lorençon E, et al. Graphene-based nanomaterials: biological and medical applications and toxicity. Nanomed., 2015; 10(15): 2423-2450.
- 43. Phiri J, Gane P, Maloney TC. General overview of graphene: Production, properties and application in polymer composites. Mater. Sci. Eng., B, 2017; 215: 9-28.
- 44. Pinto AM, Gonçalves IC, Magalhães FD. Graphene-based materials biocompatibility: A review. Colloids Surf., B, 2013; 111: 188-202.
- 45. Sanchez VC, Jachak A, Hurt RH, Kane AB. Biological interactions of graphene-family nanomaterials: an interdisciplinary review. Chem. Res. Toxicol., 2011; 25(1): 15-34.
- 46. Papageorgiou DG, Kinloch IA, Young RJ. Mechanical properties of graphene and graphene-based nanocomposites. Prog. Mater. Sci., 2017; 90: 75-127.
- 47. Choi W, Lahiri I, Seelaboyina R, Kang YS. Synthesis of graphene and its applications: a review. Crit. Rev. Solid State Mater. Sci., 2010; 35(1): 52-71.
- 48. Li K, Liu W, Ni Y, Li D, Lin D, Su Z, et al. Technical synthesis and biomedical applications of graphene quantum dots. J.

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Mater. Chem., B, 2017; 5(25): 4811-4826.

- 49. Avouris P, Dimitrakopoulos C. Graphene: synthesis and applications. Mater. today, 2012; 15(3): 86-97.
- 50. Dong L, Yang J, Chhowalla M, Loh KP. Synthesis and reduction of large sized graphene oxide sheets. Chem. Soc. Rev., 2017; 46(23): 7306-7316.
- 51. Kim KS, Zhao Y, Jang H, Lee SY, Kim JM, Kim KS, et al. Largescale pattern growth of graphene films for stretchable transparent electrodes. Nature, 2009; 457(7230): 706-710.
- 52. Smovzh DV, Kostogrud IA, Sakhapov SZ, Zaikovskii AV, Novopashin SA. The synthesis of few-layered graphene by the arc discharge sputtering of a Si-C electrode. Carbon, 2017; 112: 97-102.
- 53. De Heer WA, Berger C, Ruan M, Sprinkle M, Li X, Hu Y, et al. Large area and structured epitaxial graphene produced by confinement controlled sublimation of silicon carbide. Proc. Natl. Acad. Sci., 2011; 108(41): 16900-16905.
- 54. Park S, An J, Jung I, Piner RD, An SJ, Li X, et al. Colloidal suspensions of highly reduced graphene oxide in a wide variety of organic solvents. Nano Lett., 2009; 9(4): 1593- 1597.
- 55. Zhou C, Jiang W, Via BK. Facile synthesis of soluble graphene quantum dots and its improved property in detecting heavy metal ions. Colloids Surf., B, 2014; 118: 72-76.
- 56. Pan D, Zhang J, Li Z, Wu M. Hydrothermal route for cutting graphene sheets into blue‐luminescent graphene quantum dots. Adv. Mater., 2010; 22(6): 734-738.
- 57. Tian R, Zhong S, Wu J, Jiang W, Shen Y, Wang T. Solvothermal method to prepare graphene quantum dots by hydrogen peroxide. Opt. Mater., 2016; 60: 204-208.
- 58. Chen W, Lv G, Hu W, Li D, Chen S, Dai Z. Synthesis and applications of graphene quantum dots: a review. Nanotechnol. Rev., 2018; 7(2): 157–185.
- 59. Zhang C, Cui Y, Song L, Liu X, Hu Z. Microwave assisted onepot synthesis of graphene quantum dots as highly sensitive fluorescent probes for detection of iron ions and pH value. Talanta, 2016; 150: 54-60.
- 60. Shinde DB, Pillai VK. Electrochemical preparation of luminescent graphene quantum dots from multiwalled carbon nanotubes. Chem. Eur. J., 2012; 18(39): 12522- 12528.
- 61. Dong Y, Shao J, Chen C, Li H, Wang R, Chi Y, et al. Blue luminescent graphene quantum dots and graphene oxide prepared by tuning the carbonization degree of citric acid. Carbon, 2012; 50(12): 4738-4743.
- 62. Li J, Wang G, Zhu H, Zhang M, Zheng X, Di Z, et al. Antibacterial activity of large-area monolayer graphene film manipulated by charge transfer. Sci. Rep., 2014; 4: 4359.
- 63. Akhavan O, Choobtashani M, Ghaderi E. Protein degradation and RNA efflux of viruses photocatalyzed by graphene–tungsten oxide composite under visible light irradiation. The J. Phys. Chem., C, 2012; 116(17): 9653- 9659.
- 64. Santhosh C, Kollu P, Doshi S, Sharma M, Bahadur D, Vanchinathan MT, et al. Adsorption, photodegradation and antibacterial study of graphene–Fe₃O₄ nanocomposite for multipurpose water purification application. RSC Adv., 2014; 4(54): 28300-28308.
- 65. Hui L, Piao J-G, Auletta J, Hu K, Zhu Y, Meyer T, et al. Availability of the basal planes of graphene oxide determines whether it is antibacterial. ACS Appl. Mater.

Interfaces, 2014; 6(15): 13183-13190.

- 66. Liu M, Zhang Q, Zhao H, Chen S, Yu H, Zhang Y, et al. Controllable oxidative DNA cleavage-dependent regulation of graphene/DNA interaction. Chem. Commun., 2011; 47(14): 4084-4086.
- 67. Zhang Y, Ali SF, Dervishi E, Xu Y, Li Z, Casciano D, et al. Cytotoxicity effects of graphene and single-wall carbon nanotubes in neural phaeochromocytoma-derived PC12 cells. ACS Nano, 2010; 4(6): 3181-3186.
- 68. Chang Y, Yang S-T, Liu J-H, Dong E, Wang Y, Cao A, et al. In vitro toxicity evaluation of graphene oxide on A549 cells. Toxicol. Lett., 2011; 200(3): 201-210.
- 69. Gurunathan S, Han JW, Dayem AA, Eppakayala V, Kim J-H. Oxidative stress-mediated antibacterial activity of graphene oxide and reduced graphene oxide in Pseudomonas aeruginosa. Int. J. Nanomed., 2012; 7: 5901- 5914.
- 70. Yang HH, McCreery RL. Elucidation of the Mechanism of Dioxygen Reduction on Metal‐Free Carbon Electrodes. J. Electrochem. Soc., 2000; 147(9): 3420-3428.
- 71. Zhao Y, Chen W-f, Yuan C-f, Zhu Z-y, Yan L-f. Hydrogenated graphene as metal-free catalyst for Fenton-like reaction. Chin. J. Chem. Phys., 2012; 25(3): 335.
- 72. Chen J, Peng H, Wang X, Shao F, Yuan Z, Han H. Graphene oxide exhibits broad-spectrum antimicrobial activity against bacterial phytopathogens and fungal conidia by intertwining and membrane perturbation. Nanoscale, 2014; 6(3): 1879-1889.
- 73. Titov AV, Král P, Pearson R. Sandwiched graphene− membrane superstructures. ACS Nano, 2009; 4(1): 229- 234.
- 74. Tu Y, Lv M, Xiu P, Huynh T, Zhang M, Castelli M, et al. Destructive extraction of phospholipids from Escherichia coli membranes by graphene nanosheets. Nat. Nanotechnol., 2013; 8(8): 594–601.
- 75. Zhang L, Jiang Y, Ding Y, Povey M, York D. Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids). J. Nanopart. Res., 2007; 9(3): 479-489.
- 76. Liu S, Hu M, Zeng TH, Wu R, Jiang R, Wei J, et al. Lateral dimension-dependent antibacterial activity of graphene oxide sheets. Langmuir, 2012; 28(33): 12364-12372.
- 77. Sawangphruk M, Srimuk P, Chiochan P, Sangsri T, Siwayaprahm P. Synthesis and antifungal activity of reduced graphene oxide nanosheets. Carbon, 2012; 50(14): 5156-5161.
- 78. Khan MS, Abdelhamid HN, Wu H-F. Near infrared (NIR) laser mediated surface activation of graphene oxide nanoflakes for efficient antibacterial, antifungal and wound healing treatment. Colloids Surf., B, 2015; 127: 281-291.
- 79. Zhu S, Luo F, Zhu B, Wang G-X. Toxicological effects of graphene oxide on Saccharomyces cerevisiae. Toxicol. Res., 2017; 6(4): 535-543.
- 80. Xie J, Ming Z, Li H, Yang H, Yu B, Wu R, et al. Toxicity of graphene oxide to white rot fungus Phanerochaete chrysosporium. Chemosphere, 2016; 151: 324-331.
- 81. Wang X, Liu X, Chen J, Han H, Yuan Z. Evaluation and mechanism of antifungal effects of carbon nanomaterials in controlling plant fungal pathogen. Carbon, 2014; 68: 798-806.
- 82. Gosheger G, Hardes J, Ahrens H, Streitburger A, Buerger H, Erren M, et al. Silver-coated megaendoprostheses in a rabbit model—an analysis of the infection rate and

toxicological side effects. Biomaterials, 2004; 25(24): 5547- 5556.

- 83. Sondi I, Salopek-Sondi B. Silver nanoparticles as antimicrobial agent: a case study on E. coli as a model for Gram-negative bacteria. J. Colloid Interface Sci., 2004; 275(1): 177-182.
- 84. Li C, Wang X, Chen F, Zhang C, Zhi X, Wang K, et al. The antifungal activity of graphene oxide–silver nanocomposites. Biomaterials, 2013; 34(15): 3882-3890.
- 85. Chen J, Sun L, Cheng Y, Lu Z, Shao K, Li T, et al. Graphene oxide-silver nanocomposite: novel agricultural antifungal agent against fusarium graminearum for crop disease prevention. ACS Appl. Mater. Int., 2016; 8(36): 24057- 24070.
- 86. Karimi L, Yazdanshenas ME, Khajavi R, Rashidi A, Mirjalili M. Using graphene/TiO 2 nanocomposite as a new route for preparation of electroconductive, self-cleaning, antibacterial and antifungal cotton fabric without toxicity. Cellulose, 2014; 21(5): 3813-3827.
- 87. Li G, Zhao H, Hong J, Quan K, Yuan Q, Wang X. Antifungal graphene oxide-borneol composite. Colloids Surf., B, 2017; 160: 220-227.
- 88. Organization WH. The world health report 2002: reducing risks, promoting healthy life: World Health Organization; 2002.
- 89. Van Boeckel TP, Gandra S, Ashok A, Caudron Q, Grenfell BT, Levin SA, et al. Global antibiotic consumption 2000 to 2010: an analysis of national pharmaceutical sales data. Lancet Infect. Dis., 2014; 14(8): 742-750.
- 90. Lesprit P, Landelle C, Brun-Buisson C. Clinical impact of unsolicited post-prescription antibiotic review in surgical and medical wards: a randomized controlled trial. Clin. Microbiol. Infect., 2013; 19(2): E91-E97.
- 91. He J, Zhu X, Qi Z, Wang C, Mao X, Zhu C, et al. Killing dental pathogens using antibacterial graphene oxide. ACS Appl. Mater. Inter., 2015; 7(9): 5605-5611.
- 92. Veerapandian M, Zhang L, Krishnamoorthy K, Yun K. Surface activation of graphene oxide nanosheets by ultraviolet irradiation for highly efficient anti-bacterials. Nanotechnology, 2013; 24(39): 395706.
- 93. Gurunathan S, Han JW, Dayem AA, Eppakayala V, Park M-R, Kwon D-N, et al. Antibacterial activity of dithiothreitol reduced graphene oxide. J. Ind. Eng. Chem., 2013; 19(4): 1280-1288.
- 94. Kurantowicz N, Sawosz E, Jaworski S, Kutwin M, Strojny B, Wierzbicki M, et al. Interaction of graphene family materials with Listeria monocytogenes and Salmonella enterica. Nanoscale Res. Lett., 2015; 10(1): 23.
- 95. Wu X, Tan S, Xing Y, Pu Q, Wu M, Zhao JX. Graphene oxide as an efficient antimicrobial nanomaterial for eradicating multi-drug resistant bacteria in vitro and in vivo. Colloids Surf., B, 2017; 157: 1-9.
- 96. Das MR, Sarma RK, Saikia R, Kale VS, Shelke MV, Sengupta P. Synthesis of silver nanoparticles in an aqueous suspension of graphene oxide sheets and its antimicrobial activity. Colloids Surf. B., 2011; 83(1): 16-22.
- 97. Ruiz ON, Fernando KS, Wang B, Brown NA, Luo PG, McNamara ND, et al. Graphene oxide: a nonspecific enhancer of cellular growth. ACS Nano, 2011; 5(10): 8100-8107.
- 98. Díez-Pascual AM, Díez-Vicente AL. Poly (propylene fumarate)/polyethylene glycol-modified graphene oxide nanocomposites for tissue engineering. ACS Appl. Mater. Interfaces, 2016; 8(28): 17902-17914.
- 99. Liu S, Zeng TH, Hofmann M, Burcombe E, Wei J, Jiang R, et al. Antibacterial activity of graphite, graphite oxide, graphene oxide, and reduced graphene oxide: membrane and oxidative stress. ACS Nano, 2011; 5(9): 6971-6980.
- 100. Chen J, Wang X, Han H. A new function of graphene oxide emerges: inactivating phytopathogenic bacterium Xanthomonas oryzae pv. Oryzae. J. Nanopart. Res., 2013; 15(5): 1658.
- 101. Krishnamoorthy K, Veerapandian M, Zhang L-H, Yun K, Kim SJ. Antibacterial efficiency of graphene nanosheets against pathogenic bacteria via lipid peroxidation. J. Phys. Chem., C, 2012; 116(32): 17280-17287.
- 102. Wang Y-W, Cao A, Jiang Y, Zhang X, Liu J-H, Liu Y, et al. Superior antibacterial activity of zinc oxide/graphene oxide composites originating from high zinc concentration localized around bacteria. ACS Appl. Mater. Interfaces, 2014; 6(4): 2791-2798.
- 103. Ghosh S, Das A. Modified titanium oxide $(TIO₂)$ nanocomposites and its array of applications: a review. Toxicol. Environ. Chem., 2015; 97(5): 491-514.
- 104. Jin J, Zhang L, Shi M, Zhang Y, Wang Q. Ti-GO-Ag nanocomposite: the effect of content level on the antimicrobial activity and cytotoxicity. Int. J. Nanomed., 2017; 12: 4209–4224.
- 105. Chang Y-N, Zhang M, Xia L, Zhang J, Xing G. The toxic effects and mechanisms of CuO and ZnO nanoparticles. Materials, 2012; 5(12): 2850-2871.
- 106. Yang K, Li Y, Tan X, Peng R, Liu Z. Behavior and toxicity of graphene and its functionalized derivatives in biological systems. Small, 2013; 9(9‐10): 1492-1503.
- 107. Xu W-P, Zhang L-C, Li J-P, Lu Y, Li H-H, Ma Y-N, et al. Facile synthesis of silver@ graphene oxide nanocomposites and their enhanced antibacterial properties. J. Mater. Chem., 2011; 21(12): 4593-4597.
- 108. Liu L, Liu J, Wang Y, Yan X, Sun DD. Facile synthesis of monodispersed silver nanoparticles on graphene oxide sheets with enhanced antibacterial activity. New J. Chem., 2011; 35(7): 1418-1423.
- 109. Ma J, Zhang J, Xiong Z, Yong Y, Zhao X. Preparation, characterization and antibacterial properties of silvermodified graphene oxide. J. Mater. Chem., 2011; 21(10): 3350-3352.
- 110. Chook SW, Chia CH, Zakaria S, Ayob MK, Chee KL, Huang NM, et al. Antibacterial performance of Ag nanoparticles and AgGO nanocomposites prepared via rapid microwaveassisted synthesis method. Nanoscale Res. Lett., 2012; 7(1): 541.
- 111. Rojas-Andrade MD, Chata G, Rouholiman D, Liu J, Saltikov C, Chen S. Antibacterial mechanisms of graphene-based composite nanomaterials. Nanoscale, 2017; 9(3): 994- 1006.
- 112. Santos CM, Mangadlao J, Ahmed F, Leon A, Advincula RC, Rodrigues DF. Graphene nanocomposite for biomedical applications: fabrication, antimicrobial and cytotoxic investigations. Nanotechnology, 2012; 23(39): 395101.